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A-109 GRIDLEY'S METHOD FOR ENDAMOEBA HISTOSLYTICA

FIXATION: **10% Buffered Neutral Formalin** (F-113).

TECHNIQUE: Cut paraffin sections at 6 microns

STAINING PROCEDURE:

1. Deparaffinize and hydrate to distilled water. Include a control slide with all samples.
2. Stain in **Harris Hematoxylin** (A-109-1), 10 min. or Weigert's Iron Hematoxylin, 3 min.

*To prepare Weigert's Iron Hematoxylin, mix equal parts just before use:
Weigert's Iron Hematoxylin Sol'n A (A-109-1A) and
Weigert's Iron Hematoxylin Sol'n B (A-109-1B).

3. Wash in running water for several minutes, differentiate each slide in **Acid Alcohol, 1%** (A-109-4) and then wash again.
4. Blue each slide in **Ammonia Water** (A-109-5), wash in running water.
5. Stain in **Aniline-Eosin** (A-109-2), 5 minutes. Rinse well in distilled water. Slides should appear deep rose.
6. Counterstain in **Naphthol Green B** (A-109-3), 5 minutes.
7. Under a microscope, differentiate the slides in 95% alcohol (two changes) until the erythrocytes remain a deep rose.
8. Dehydrate through two changes absolute alcohol and clear in two changes **Xylene** (C-120).
9. Mount with **Permount** (M-18).

RESULTS:

Erythrocytes (ingested)	rose
Connective tissue	green
Amoebae	blue-green
Nuclei (amoebic)	darker blue-green

REFERENCES:

Gridley, M.F.: Am. J. Clin., Pathol. 24:243 (1954).

Luna, L.G.: Histologic Staining Methods, 3rd ed.: New York: McGraw-Hill Book Co., c. 1968, p. 228.