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A-141 LENDRUM'S METHOD FOR INCLUSION BODIES

Fixation: **10% Buffered Neutral Formalin** (F-113) or **Zenker's** (F-155)

Sections: Cut paraffin at 6 microns

Staining:

1. Deparaffinize and hydrate to distilled water.
2. If Zenker's fixed, place in **Lugol's Iodine** (A-141-4) or **Gram's Iodine** (A-141-4A) for 15 minutes, then rinse in water. (If formalin fixed, was use proceed with step 4).
3. Place in **Sodium Thiosulfate 5%** (A-141-5), for 3 minutes and wash in running water for 10 minutes or longer.
4. Stain in **Mayer's Hematoxylin** (A-141-l), for 5-10 minutes and then wash in running water until sections become blue (approx. 15 minutes).
5. Stain in **Phloxine-Calcium Chloride** (A-141-2), for 30 minutes. Rinse briefly in distilled water and drain slides well.
6. Differentiate with the **Tartrazine-Cellosolve Solution** (A-141-3), by flooding the slide. The differentiation time can vary from a few minutes to several hours and, therefore, the differentiation should be followed with a microscope. Differentiation should proceed until the inclusion bodies are a bright red and before they begin to fade.
7. Wash briefly in water to remove enough Tartrazine to produce a pleasing yellow background.
8. Dehydrate through 60%, 95%, and absolute alcohol, clear in 2 changes of Xylene, and mount with Permount (M-18)

Staining Results:

Inclusion Bodies	Bright Red
Collagen and background	Yellow
Nuclei	Blue

REFERENCES:

Lendrum, A.C., J. Path. Bact., 59:399-404, 1947
Clark, 0., (ed), Staining Procedures, Williams and Wilkins Co., Baltimore, 3rd. edit., c 1973, pg 341
Luna, L.G. (ed) , Manual of Histological Staining Methods of the Armed Forces Institute of Pathology, McGraw-Hill, NY, 3rd. ed. , a 1968, p 234