A-141  LENDRUM’S METHOD FOR INCLUSION BODIES

Fixation: 10% Buffered Neutral Formalin (F-113) or Zenker’s (F-155)

Sections: Cut paraffin at 6 microns

Staining:
1. Deparaffinize and hydrate to distilled water.
2. If Zenker’s fixed, place in Lugol’s Iodine (A-141-4) or Gram’s Iodine (A-141-4A) for 15 minutes, then rinse in water. (If formalin fixed, was use proceed with step 4).
3. Place in Sodium Thiosulfate 5% (A-141-5), for 3 minutes and wash in running water for 10 minutes or longer.
4. Stain in Mayer’s Hematoxylin (A-141-l), for 5-10 minutes and then wash in running water until sections become blue (approx. 15 minutes).
5. Stain in Phloxine-Calcium Chloride (A-141-2), for 30 minutes. Rinse briefly in distilled water and drain slides well.
6. Differentiate with the Tartrazine-Cellosolve Solution (A-141-3), by flooding the slide. The differentiation time can vary from a few minutes to several hours and, therefore, the differentiation should be followed with a microscope. Differentiation should proceed until the inclusion bodies are a bright red and before they begin to fade.
7. Wash briefly in water to remove enough Tartrazine to produce a pleasing yellow background.
8. Dehydrate through 60%, 95%, and absolute alcohol, clear in 2 changes of Xylene, and mount with Permount (M-18)

Staining Results:
- Inclusion Bodies: Bright Red
- Collagen and background: Yellow
- Nuclei: Blue

REFERENCES:
- Clark, 0., (ed), Staining Procedures, Williams and Wilkins Co., Baltimore, 3rd. edit., c 1973, pg 341

January 26, 2018