

## **B-169     TURNBULL BLUE METHOD FOR HEMOSIDERIN**

**FIXATION:** **Buffered Neutral Formalin, 10%** (F—113) or absolute alcohol.

**SECTIONS:** Cut paraffin @ 6 microns.

**STAINING:**

1. Deparaffinize and hydrate to distilled water.
2. **Ammonium Sulfide** (B-169-1) for 4 hours. Rinse thoroughly in distilled water.  
NOTE: Loss of sections sometimes occurs in Ammonium Sulfide. Dilution of one part Ammonium Sulfide with 3 parts of 95% alcohol is a way to avoid this problem.
3. Stain in working\* Hydrochloric Acid — Potassium Ferricyanide solution for 20 minutes. Rinse thoroughly in distilled water.  
\*Prepare working stain by mixing equal volumes of **Hydrochloric Acid, 1%** (B-169-2) and Potassium **Ferricyanide, 20%** (B-169-3), just before use.
4. Counterstain for 5 minutes in **Nuclear Fast Red Solution** (B-169-4). Rinse in distilled water.
5. Dehydrate in 95% alcohol, absolute alcohol, and clear in Xylene, two changes each.
6. Mount with **Permount** (M-18).

**STAINING RESULTS:**

Hemosiderin	blue
Nuclei	red

**REFERENCES:**

Mallory, F 3., Pathological Technique, Hafner Publ. Co., NY, 1961, p 138

Luna,L.G.,(ed.), Manual of Histologic Staining Methods of the AFIP, 3<sup>rd</sup> edition, McGraw-Hill, N.Y., p.178, c. 1968