

## E-306 LAQUEUR'S METHOD FOR ALCOHOLIC HYALIN

**Fixation:** 10% Buffered Neutral Formalin (F-113) or Zenker's

**Sections:** Paraffin at 6 microns

**Staining:**

1. Deparaffinize and hydrate to distilled water.
2. Stain in **Mayer's Hematoxylin** (E-306-1) for 15 minutes. Wash in running water for 15 minutes.
3. Flood slides with **Acid Fuchsin - Aniline Solution** (E-306-2), *shake bottle well before using solution will separate upon standing*, and heat gently until aniline fumes escapes; *view against a dark background*, Let stand for 5 minutes. Wash in water, 3 dips.
4. Differentiate in **Alcoholic Picric Acid** (E-306-5) until only hyalin and red cells remain red, and collagen is faint gray or unstained. Wash in running water for 3 minutes.
5. Mordant in 1% Phosphomolybdic Acid (E-306-4) for 4 to 18 hours.
6. Counterstain in Light Green Solution (E-306-3) for 1 hour. Wash in running water for 1 minute.
7. fibers are seen as distinct fibrils.
8. Dehydrate in 95% alcohol, absolute alcohol, and clear in Xylene, two changes each.
9. Mount with **Permount** (M-18).

**Results:**

Mallory Bodies	Bright red
Erythrocytes	Red
Cytoplasm	Pale brown
Bile pigment	Green
Proteinaceous material seen occasionally in some liver cells	Red
Hemosiderin and lipofichsin	Unstained

**References:**

Laqueur, G.L., *Amer. J. Clin. Path.*, 20:689-690, 1950.

Luna, L.G. (ed), *Manual of Histologic Staining Methods of the AFIP*, 3<sup>rd</sup> edition, McGraw-Hill, NY, c 1968, p 160.

Lillie, R.D., Fullmer, H.M., *Histopathologic Technic and Practical Histochemistry* 4<sup>th</sup> ed., McGraw-Hill, N.Y., 1976, pp670-671.

Clark, G., (ed.) *Staining Procedures*, 3<sup>rd</sup> edition, Williams & Wilkins, Baltimore, p. 52 c. 1973