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F-360 , MALLORY'S ANILINE BLUE COLLAGEN STAIN

Fixation: Zenker's (F-155A +F-155B)
Sections: Paraffin@ 6 microns

Staining:

1. Deparaffinize and hydrate to distilled water.
2. Place in **Lugol's Iodine** (F-360-3) for 15 minutes. Rinse in tap water.
3. Place in **Sodium Thiosulfate, 5%** (F-350-4) for 3 minutes. Wash in tap water for 10 minutes or longer. Rinse in distilled water.
4. **Acid Fuchsin Solution** (F-360-1) for 1 to 5 minutes. *Standing with Acid Fuchsin Solution may be omitted if it is desired that collagen fibers stand out more sharply.*
5. Transfer directly to **Anilne Blue-Orange G Solution** (F-360-2) for 30 to 60 minutes or longer.
6. Dehydrate in 95% alcohol, absolute alcohol, and clear in **Xylene** (C-120), two changes each.
7. Mount with **Permount** (M-18).
   Note: For celloidin sections shorten the staining time, decolorize, and dehydrate in 95% alcohol and clear by the blotting-paper-xylene method or in terpineol. Mount in Balsam.

Results:

Nuclei	Red
Fibroglia	Red
Myoglia	Red
Neuroglia fibrils	Red
Axis cylinders	Red
Fibrin	Red
Nucleoli	Red
Collagenous fibrils	Blue
Ground substances of cartilage, bone, mucus and amyloid	Varying shades of Blue
Blood Corpuscles	Yellow
Myelin	Yellow
Elastic fibrils	Pale Pink or Pale yellow or Unstained

References:

Clark, g.: Staining Procedures, Williams and Wilkins Co., Baltimore, 3rd Ed., c. 1973, p. 54

AFIP Manual of Histological Staining Methods, 3rd Ed., Ed. L. Luna: New York: McGraw-Hill Publications, c. 1968. P. 75

Mallory, F.B., J. Exper. Med., 5:15-20, 1900.

Mallory, F.B., Pathological Technique, W.B. Saunders Co., Phila., 1938.