

F-370 WEIGERT'S RESORCIN FUCHSIN

FIXATION: Any common reagent may be used.

SECTION: Cut paraffin sections at 6 microns

STAINING PROCEDURE:

1. Deparaffinize and hydrate to distilled water.
2. Stain in **Resorcin-Fuchsin Solution** (F-370-1), for 1-12 hours depending upon the depth of stain required for the elastic fibers. (Nuclei may be stained in Weigerts' Iron Hematoxylin prior to the **Resorcin Fuchsin** rather than after as is described here.)
 - *To prepare Weigerts' Hematoxylin solution
 - *Prepare fresh just before use.
 - Mix equal parts of **Weigert's Iron Hematoxylin A** (F-370-4A), with **Weigert's Iron Hematoxylin B** (F-370-4B).
3. Clear the stain in 95% Alcohol. Acid Alcohol may be used to differentiate any slide that has stained diffusely, wash in tap water.
4. Stain slides in Weigert's Iron Hamatoxylin (see above) or other Alum Hematoxylin (**Delafield's Hematoxylin**), (F-370-3A) and then counterstain in **Van Gieson's Solution**, (F-370-3B). **Orth's Carmine Stain**, (F-370-2) may be used to stain the nuclei prior to the Resorcin-Fuchsin for a resulting reddish dust and where no other counterstain is desired.
 - Water will remove excess Van Gieson's Stain from slides.
5. Differentiate and dehydrate the slides in two changes each of 95% Alcohol and Absolute Alcohol. Clear the slides in two changes of Xylene.
6. Mount

RESULTS:

Elastic Tissues.....dark blue
Nuclei (red if Orth's Carmine used).....dark blue
Collagen.....red-pink
Miscellaneous Tissue Elements.....yellow
(with Van Gieson's stain)

REFERENCES:

Mallory, F.B.: **Pathological Technique**; New York: Hafner Publishing Co., c. 1961, p. 168.

Clark, G.: **Staining Procedures**; Baltimore: Williams and Wilkins Co., 3rd ed., c. 1973, p. 60

Luna, L.G.: **AFIP Histologic Staining Methods**; New York: McGraw-Hill Book Co., 3rd ed., c. 1968, p. 80