

**K-693**

**SHEEHAN'S METHOD FOR  
BODIAN'S PROTARGOL**

**NO COUNTERSTAINS USED!**

**Fixation:** Formalin, 10%

**Sections:** Paraffin @ 6-8 microns

**Staining:**

1. Deparaffinize and hydrate to distilled water.
2. Place slides in **Protargol Solution** (K-693-1) containing 4-6 gm clean **Copper Shot** (K-693-2) per 100 ml of solution. Let stand at 37°C for 12-48 hours.  
Staining is complete when sections are golden brown.
3. Wash in distilled water.
4. Reduce in **Hydroquinone Reducing Solution** (K-693-5) for 10 minutes. Wash thoroughly in distilled water.
5. Tone in **Acetic Gold Chloride, 1%** (K-693-3A) for 5-10 minutes.  
*To prepare Acetic Gold Chloride mix:*  

100 ml	Gold Chloride, 1% (K-693-3)
3 drops	Glacial Acetic Acid
6. Wash in distilled water.
7. If sections do not have a light purple color, place in **Oxalic Acid, 2%** (K-693-6) until sections have a definite purplish tinge (5-10 minutes). Wash in distilled water.
8. Remove the residual silver salts with **Sodium Thiosulfate, 5%** (K-693-7) for 5-10 minutes.
9. Wash thoroughly in distilled water.
10. Dehydrate rapidly in 95% alcohol, absolute alcohol and clear in Xylene (C-120), two changes each.
11. Mount.

**Results:** Myelinated fibers, the finest nonmyelinated fibers of central and peripheral nervous system, and neurofibrils- **BLACK**

**References:** Sheehan and Hrapchak, Theory and Practice of Histotechnology. St. Louis, The Mosby Company 1980 p. 256.