

K-699, HOLZER'S METHOD FOR GLIAL FIBERS

Fixation: Buffered Neutral Formalin, 10% (F-113) or Formal Alcohol

Sections: Paraffin @ 6 microns.

Staining:

1. Deparaffinize and hydrate to distilled water. Carry slides through one at a time.
2. **Phosphomolybdic Acid-Alcohol Working Solution** for 3 minutes. Drain.
To prepare working solution mix:
50ml of Phosphomolybdic Acid, 0.5% (K-699-4)
100ml of 95% Alcohol
Prepare fresh.
3. **Alcohol-Chloroform Solution** (K-699-6) until section becomes translucent.
4. Place slide on staining rack and flood section with **Crystal Violet Solution** (K-699-1) for 30 seconds.
Blot dry.
5. Flood section with **Potassium Bromide Solution** (K-699-2) for one minute. Blot dry.
6. **Differentiating Solution** (K-699-3) for 30 seconds. If over-differentiated, restain.
7. Rinse in xylene, several changes. Check with microscope after first xylene.
8. Mount with **Permount** (M-18).

Staining Results:

Glial fibers	Deep violet
Background	Pale Violet

Note: If a Crystal Violet precipitate forms on the slide, it may be removed with straight aniline.

References:

AFIP, Manual of Histological Staining Methods, 3rd ed., Ed. L. Luna: New York: McGraw Hill Publications, c. 1968, p. 200.

Holzer, W., Z. ges. Neurol. Psychiat., 69:354, 1921.