

## **L-756, MAYER'S HEMATOXYLIN**

### **Staining:**

1. Deparaffinize and hydrate to distilled water.
2. If sections are **Zenker**-Fixed , remove mercuric Chloride crystals with Iodine (Gram's or Lugol's) and clear with **Sodium Thiosulfate 5%** (A-131-5).
3. **Mayer's Hematoxylin** (L-756-1A) for 15 minutes or **Mayer's Acid Hemalum** (L-756-1B) for 5 minutes.  
Slides may be left in the Hematoxylin for hours without overstaining.
4. Wash in running tap water for 20 minutes. Note: Slides stained with Mayer's Hematoxylin will fade in 1 to 3 years time unless washed for a minimum of 20 minutes in running water.
5. Counterstain with **Eosin Y** (L-756-2).
6. Dehydrate in 95% alcohol and absolute alcohol, two changes, two minutes each. Check under microscope to see that excess Eosin is removed.
7. Clear in **Xylene** (C-120) two changes two minutes each. Mount.

NOTE: **Mayer's Hematoxylin** will sometimes stain adhesives used to attach sections onto slides. This will give the slides a dark appearance, but will not affect nuclear staining. Slides can be cleaned with a solution of 10% Acetic Acid in 95% alcohol, after Mayer's Hematoxylin - step 3.

### **Stain Results:**

Nuclei	Blue
Cytoplasm	Pink

### **REFERENCES:**

AFIP Manual of Histological Staining Methods, 3<sup>rd</sup> ed., ED. L. Luna: New York: McGraw Hill Publications, c. 1968, p. 37.