L-756, MAYER'S HEMATOXYLIN

Staining:

1. Deparaffinize and hydrate to distilled water.

2. If sections are Zenker-Fixed, remove mercuric Chloride crystals with Iodine (Gram's or Lugol's) and clear with Sodium Thiosulfate 5% (A-131-5).

3. **Mayer's Hematoxylin** (L-756-1A) for 15 minutes or **Mayer's Acid Hemalum** (L-756-1B) for 5 minutes.
   Slides may be left in the Hematoxylin for hours without overstaining.

4. Wash in running tap water for 20 minutes. Note: Slides stained with Mayer's Hematoxylin will fade in 1 to 3 years time unless washed for a minimum of 20 minutes in running water.

5. Counterstain with Eosin Y (L-756-2).

6. Dehydrate in 95% alcohol and absolute alcohol, two changes, two minutes each. Check under microscope to see that excess Eosin is removed.

7. Clear in Xylene (C-120) two changes two minutes each. Mount.

**NOTE:** Mayer's Hematoxylin will sometimes stain adhesives used to attach sections onto slides. This will give the slides a dark appearance, but will not affect nuclear staining. Slides can be cleaned with a solution of 10% Acetic Acid in 95% alcohol, after Mayer's Hematoxylin - step 3.

**Stain Results:**

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<tbody>
<tr>
<td>Nuclei</td>
<td>Blue</td>
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<tr>
<td>Cytoplasm</td>
<td>Pink</td>
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**REFERENCES:**